Maltodextrins Based Solid Membranes for the Enantioanalysis of L-Cysteine

Raluca-Ioana Stefan-van Staden^{*} and Luxolo Holo

Laboratory of Electrochemistry and PATLAB Bucharest, National Institute of Research for Electrochemistry and Condensed Matter, 202 Splaiul Independentei Str., 060021, Bucharest, Romania

Abstract: Three enantioselective membranes based on maltodextrins with different values of dextrose equivalent (DE) were proposed for the enantioanalysis of L-cysteine. The membranes were used for the design of potentiometric sensors. The slopes of the sensors were near-Nernstian (higher than 58.00mV/decade of concentration) with limits of detection of magnitude order of 10⁻¹¹ and 10⁻¹² mol/L. The surfaces of the membranes were stable for more than 6 months of continuous use. They can be renewed by polishing on alumina paper.

Keywords: L-cysteine, solid enantioselective membrane, maltodextrin, enantioselective, potentiometric membrane electrode.

1. INTRODUCTION

L-Cysteine (L-Cys) is a sulphur-containing amino acid R-SH and one of the twenty basic proteins [1]. It can be used as a prospective radiation protector and cancer indicator [2-5]. The electrochemical study of cysteine has been reported [6-9]. The development of chemically modified amperometric electrodes for detection of cysteine is a growing field [10-16]. Fluorescence was also employed for the assay of cysteine [17].

Chiral recognition is an area of considerable research interest due to its importance in biological, chemical and pharmaceutical sciences [18]. Techniques, which are intensively used for chiral recognition are: chromatography, capillary zone electrophoresis, mass spectrometry and electrochemistry. Utilization of electrochemical techniques will make the enantioanalysis highly reliable [19].

Maltodextrins represents a class of very powerful chiral selectors [20-22]. Variations in DE values result in maltodextrins with varying physico-chemical properties: solubility, hydroscopicity, osmolality and their effectiveness to reduce the freezing point increase with increasing DE, while viscosity, cohesiveness and coarse-crystal prevention increase as DE decreases [23, 24]. Maltodextrins were used as chiral selectors for enantiomeric separations by capillary zone electrophoresis [21, 22, 25-29], and they were also used for the design of enantioselective, potentiometric membrane electrodes [19, 30-33]. Although, the HPLC

(standard method) method, and fluorescence based method are highly used for biomedical analysis, they cannot always be high reliable especially for urine samples, when the complexity of the sample is very high.

This paper proposed three solid enantioselective membranes used in the design of enantioselective, potentiometric membrane electrodes (EPMEs) for the enantioanalysis of L-cysteine. The membranes were based on maltodextrins with different DE.

2. EXPERIMENTAL

2.1. Reagents and Materials

L- and D-Cysteine were bought from Sigma-Aldrich (St. Louis, MO, USA). Maltodextrins (DE 4.0-7.0, 13.0-17.0, 16.5-19.5) and graphite powder (1-2 μ m, synthetic) were bought from Aldrich (Milwaukee, WI, USA).

Deionised water was obtained using a Modulab system (Continental Water Systems, San Antonio, TX, USA). The L- and D-cysteine solutions necessarily in the characterization of the enantioselective, potentiometric membrane electrodes were prepared from standard L- and D-cysteine solutions (10⁻² mol/L), respectively, by serial dilutions. All solutions were buffered with phosphate buffer (pH 2.40, 0.1mol/L) from Merck (Darmstadt, Germany) (1:1, v/v, buffer: deionised water).

2.2. Apparatus

The potentiometric measurements were done using a system comprising a 663 VA stand (Metrohm, Herisau, Switzerland), a PGSTAT 20 (Eco Chemie,

^{*}Address correspondence to this author at the Laboratory of Electrochemistry and PATLAB Bucharest, National Institute of Research for Electrochemistry and Condensed Matter, 202 Splaiul Independentei Str., 060021, Bucharest, Romania; Tel: +40751507779; E-mail: iustinavanstaden@yahoo.com

Utretch, Netherlands), and a software version 4.9. Ag/AgCl (0.1 mol/L KCl) was used as reference electrode in the cell.

2.3. Solid Membranes and Electrodes Design

Paraffin oil was added to graphite powder in a ratio of 1:4 (w/w). A solution of maltodextrin (DE 4.0-7.0 (I), 13.0-17.0 (II), or 16.5-19.5 (III); 10^{-3} mol/L) was added to the paste in a ratio 1:1 (µL:mg). The diameter of the active area of the potentiometric, enantioselective membrane electrode was 3 mm. Ag/AgCl was used as electric contact. 0.1 mol/L KCl solution was used as internal solution.

2.4. Recommended Procedures

Direct potentiometric method was used for all measurements using solution with concentrations between 10^{-10} and 10^{-2} mol/L. The working and reference electrodes were placed in stirred standard solutions. Graphs of E(mv) versus pL-Cys were plotted and unknown concentrations were determined from the graphs.

2.4.1. Determination of L-Cysteine in Urine Samples

Urine samples were collected from different patients and buffered with phosphate buffer (pH 2.40, 0.1mol/L) (1:1, v/v, buffer:urine sample). Direct potentiometric method was used to determine L-cysteine in urine samples.

3. RESULTS AND DISCUSSION

3.1. Electrodes Response

The response characteristics of the electrodes were determined using potentiometric method (Table 1). All calibration equations had correlation coefficients of 0.9999. The limits of detection are very low – of 10^{-11} and 10^{-12} mol/L magnitude order. The response of the proposed electrodes was between 58.00 and 60.00

mV/decade of concentration, very closed with the Nernstian value. The electrodes did not respond for D-cysteine assay; 3-5mV/decade of concentration was obtained when the three sensors were used.

The proposed electrodes were highly stable and reproducible over 6 months test period (RSD value for the slopes did not exceeded 1%). The response time was 20s for concentration range $10^{-5} - 10^{-3}$ mol/L and 1min for concentrations lower than 10^{-5} mol/L.

3.2. The Effect of pH on the Response of the Electrodes

The influence of pH on the response of the proposed electrodes was investigated for solutions of 10^{-5} mol/L L-Cys at different pH values (pH 1-12). These solutions were prepared by addition of small volumes of HCl and/or NaOH solution (0.1-1 mol/L of each) to a L-Cys solution.

The plots of E (mV) versus pH (Figure 1) show that the response of the electrodes is not depending on pH, in the following ranges 2.0-5.0, 2.0-7.0, and 2.0-6.0 for the EPMEs based on maltodextrins I, II, and III, respectively.

3.3. The Selectivity of the Electrodes

Mixed solution method was used for the study of the selectivity of the proposed electrodes versus D-Cys, polyvinylpyrolidone (PVP), creatine, creatinine, Na⁺, K⁺, and Ca²⁺. The concentration of the interfering ions and L-Cys were 10^{-4} and 10^{-5} mol/L, respectively. The EPMEs based on maltodextrins were selective over PVP, creatine and creatinine and enantioselective (Table **2**). Potentiometic selectivity coefficients were calculated using the equation:

$$K_{i,j}^{pot} = \left(10^{\frac{\Delta E}{S}} - 1\right) \times \frac{a_i}{a_i^{z_i/z_j}}$$

Table 1: Response Characteristics of Enantioselective, Potentiometric Membrane Electrodes for the Assay of L-Cysteine

EPME based on maltodextrin	Slope (mV/decade of conc.)	Intercept, E° (mV)	Linear conc. range (mol/L)	Detection limit (mol/L)
I	58.5±0.2	587.5±12.2	10 ⁻¹⁰ -10 ⁻³	9.0x10 ⁻¹²
II	59.0±0.1	659.8±11.3	10 ⁻¹⁰ -10 ⁻³	5.2x10 ⁻¹²
III	59.2±0.2	600.7±12.5	10 ⁻¹⁰ -10 ⁻³	7.1x10 ⁻¹¹

All measurements were made at 25°C. All values are averages of ten determinations.

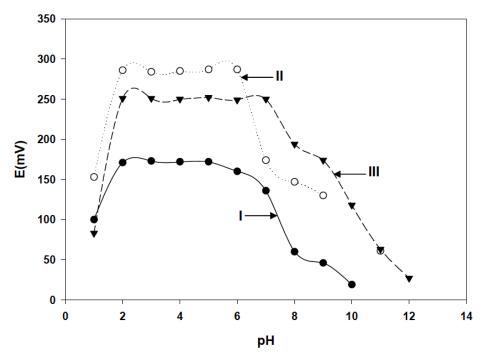


Figure 1: The influence of pH on the response of the enantioselective, potentiometric membrane electrodes (C_{L-Cys}=10⁻⁵mol/L); I - MD-I, II - MD-II and III - MD-II based EPMEs.

where ΔE is the difference between the potential recorded for mixed solution (E_{i,j}) and for the solution that contains only the main ion (E_i), $\Delta E = E_{i,j} - E_i$ (all recorded in mV); S is the slope of the electrode deducted from the equation of calibration (mV/decade of concentration); a_i is the activity of the main ion, i, a_j is the activity of interfering species, j; z_i is the charge of the main ion, i, z_j is the charge of interfering species, j.

Inorganic ions such as Na⁺, K⁺, and Ca²⁺ did not interfere with the analysis of L-Cys, because the potentiometric selectivity coefficients calculated were less than 10⁻⁴.

3.4. Analytical Applications

Recovery tests were performed first for the assay of L-cys in the presence of D-cys; different ratios between

L- and D-Cys concentrations were used. The results obtained (Table **3**) demonstrated that D-Cys did not interfere in the assay of L-Cys.

L-cysteine was reliable determined in urine samples; results are shown in Table **4**.

The average recovery of L-cysteine in urine samples was higher that 99% from the amount of L-cysteine determined using a standard method - HPLC [34]. These values were taken as reference for the validation of the proposed method.

4. CONCLUSION

Maltodextrins were excellent chiral selectors for the design of enantioselective membranes. The construction of the membranes was simple, fast, and

Table 2:	Potentiometric, Selectivity	Coefficients of the	e Electrodes Used for the	Enantioanalysis of L-Cysteine

Interfering species	$K_{i,j}^{pot}$			
(J)	EPME based on			
	MD-I	MD-II	MD-III	
D-Cys	6.0x10 ⁻³	8.1x10 ⁻⁴	4.0x10 ⁻⁴	
PVP	3.2x10 ⁻³	3.2x10 ⁻³	8.1x10 ⁻⁴	
Creatine	4.0x10 ⁻⁴	3.3x10 ⁻³	4.0x10 ⁻⁴	
Creatinine	<< 1.0x10 ⁻⁴	4.0x10 ⁻⁴	8.1x10 ⁻⁴	

All measurements were made at 25°C. All values are averages of ten determinations.

Table 3: Determination of L-Cysteine in the Presence of D-Cysteine

		L-cys, Recovery, %		
L:D mol/mol	EPME based on MD			
	I	II	III	
2:1	99.63±0.01	99.72±0.02	99.98±0.02	
1:1	99.95±0.02	99.99±0.01	99.60±0.02	
1:2	99.81±0.02	100.00±0.01	99.24±0.01	
1:4	99.90±0.01	99.97±0.02	100.00±0.01	
1:9	99.54±0.02	99.99±0.01	99.99±0.02	

All measurements were made at 25°C. All values are averages of ten determinations.

Table 4: Determination of L-Cysteine in Urine Samples

	L-cys, Recovery			
Urine sample	EPME based on M	HPLC [34],		
	I	I	111	mol/L
1	99.42±0.20	99.80±0.15	99.99±0.25	2.3x10 ⁻⁶
2	99.50±0.15	99.58±0.18	99.59±0.20	2.7x10 ⁻⁷
3	99.09±0.20	99.86±0.21	99.49±0.10	3.2x10 ⁻⁷
4	100.01±0.21	100.00±0.20	99.99±0.21	4.3x10 ⁻⁷
5	99.87±0.10	99.85±0.12	99.57±0.21	4.0x10 ⁻⁷
6	99.39±0.15	99.66±0.20	99.19±0.18	3.9x10 ⁻⁷

All measurements were made at 25°C. All values are averages of ten determinations.

reproducible. The electrodes' selectivity and enantioselectivity made them suitable for enantioanalysis of L-Cysteine in urine samples. The best maltodextrin based electrode for the enantioanalysis of L-cysteine proved to be the one based on maltodextrin III because it exhibited the best selectivity and enantioselectivity as well as the highest slope. Accordingly, this electrode is the electrode of choice for the enantioanalysis of L-cysteine.

ACKNOWLEDGEMENTS

The authors are grateful to the PNII Program Capacity, 2012-2014, Contract nr. 3ERC-like/2012 for financial support.

REFERENCES

- Voet D, Voet JG, Biochemistry, 2nd ed., New York: Wiley 1995, p. 1263.
- [2] Kulys J, Drungiliene A, Chemically modified electrodes for the determination of sulphydryl compounds. Anal. Chim. Acta 1991; 243: 287-92. <u>http://dx.doi.org/10.1016/S0003-2670(00)82572-6</u>
- [3] Townshend A Ed., Encyclopedia of Analytical Science, Vol. 3, London: Academic Press 1995, p. 1735.

- [4] Filanovsky B, Electrochemical response of new carbon electrodes bulk modified with cobalt phthalocyanine to some thiols in the presence of heptane or human urine. Anal Chim Acta 1999; 394: 91-100. <u>http://dx.doi.org/10.1016/S0003-2670(99)00035-5</u>
- [5] Nagasawa HT, Elberling JA, Roberts JC, Beta-substituted cysteines as sequestering agents for ethanol-derived acetaldehyde *in vivo*. J Med Chem 1987; 30: 1373-8. <u>http://dx.doi.org/10.1021/jm00391a018</u>
- [6] Persson B, A Chemically modified graphite electrode for electrocatalytic oxidation of reduced nicotinamide adenine dinucleotide based on a phenothiazine derivative, 3-βnaphthoyl-toulidine blue. J Electroanal Chem 1990; 287: 61-80.

http://dx.doi.org/10.1016/0022-0728(90)87160-L

- [7] Ke B. The polarographic behaviour of α -lipoic acid. Biochem Biophys Acta 1957; 25: 650-1. http://dx.doi.org/10.1016/0006-3002(57)90544-9
- [8] Zagal JH, Metallophthalocyanines as catalyst in electrochemical reactions. Coord Chem Rev 1992; 119: 89-136. http://dx.doi.org/10.1016/0010-8545(92)80031-L
- [9] Arrigan DWM, Bihan LL. A study of L-cysteine adsorption on gold via electrochemical desorption and copper (II) ion complexation. Analyst 1999; 124: 1645-9. <u>http://dx.doi.org/10.1039/a905370k</u>
- [10] Chen SM. Electrocatalytic reaction, catalytic autoxidation, and supported catalytic autoxidation of sulfur oxoanions by FeTSPP and Mn(4-TMPyP). J Electroanal Chem 1996; 407: 123-30.

http://dx.doi.org/10.1016/0022-0728(95)04464-7

Stefan-van Staden and Holo

- Zagal JH, Aguirre MJ, Parodi CG, Sturm J, Electrocatalytic [11] activity of vitamin B₁₂ adsorbed on graphite electrode for the oxidation of cysteine and glutathione and the reduction of cysteine. J Electroanal Chem 1994; 374: 215-22. http://dx.doi.org/10.1016/0022-0728(94)03365-X
- Sugawara K, Hoshi S, Akatsuka K, Shimazu K, [12] Electrochemical behaviour of cysteine at a Nafion®|cobalt(II) modified electrode. J Electroanal Chem 1996; 414: 253-6.
- Sugarawa K, Tanaka S, Taga M. Voltametric behaviour of [13] cysteine by a carbon-paste electrode containing cobalt(II) cyclohexylbutyrate. Bioelectrochem Bioenerg 1991; 26: 469-74 http://dx.doi.org/10.1016/0302-4598(91)85008-P
- [14] Halbert MK, Baldwin RP. Electrocatalytic and analytical response of cobalt phthalocyanine containing carbon paste electrodes toward sulfhydryl compounds. Anal Chem 1985; 57: 591-5. http://dx.doi.org/10.1021/ac00280a007
- [15] Mafatle TJ, Nyokong T. Electrocatalytic oxidation of cysteine by molybdenum (V) phthalocyanine complexes. J Electroanal Chem 1996; 408: 213-8. http://dx.doi.org/10.1016/0022-0728(95)04519-8
- Pang DW, Wang ZL, Electrocatalysis of metalloporphyrins: [16] Part 13. Electrocatalysis of several water-soluble porphyrins for the oxidation of some small molecules. J Electroanal Chem 1993; 358: 235-46. http://dx.doi.org/10.1016/0022-0728(93)80441-J
- Huang S, Xiao Q, Li R, Guan HL, Liu J, Liu XR, He ZK, Liu Y, [17] A simple and sensitive method for I-cysteine detection based on the fluorescence intensity increment of quantum dots. Anal Chim Acta 2009; 645: 73-8. http://dx.doi.org/10.1016/j.aca.2009.04.034
- [18] Aboul-Enein HY. Wainer IW. The Impact of Stereochemistry on Drug Development and Use, New York: Wiley 1997.
- Stefan RI, van Staden JF, Aboul-Enein HY, Electrochemical [19] Sensors in Bioanalysis, New York: Marcel Dekker 2001.
- Soini M, Stefansson M, Riekkola ML, Novotny NV. [20] Maltooligosaccharides as chiral selectors for the separation of pharmaceuticals by capillary electrophoresis. Anal Chem 1994; 66: 3477-84. http://dx.doi.org/10.1021/ac00092a028
- [21] Huslt AD, Verbeke N, Separation of the enantiomers of coumarinic anticoagulant drugs by capillary electrophoresis using maltodextrins as chiral modifiers. Chirality 1994; 6: 225-9.
- [22] Huslt AD, Verbeke N. Chiral analysis of basic drugs by oligosaccharide mediated capillary electrophoresis. J Chromatogr A 1996; 735: 283-93. http://dx.doi.org/10.1016/0021-9673(95)01356-3

DOI: http://dx.doi.org/10.6000/1929-6037.2014.03.02.3

Received on 25-04-2014

Accepted on 14-05-2014

Published on 31-05-2014

- Altshul AM, Low caloric foods-a scientific status summary by [23] the Institute of Food Technologies expert panel of food safety and nutrition. Food Technol 1989: 43: 113-20.
- [24] On the molecular characteristics, Chronakis IS, compositional properties and and structural-functional mechanism of the maltodextrins: a review, Crit Rev Food Sci. 1998: 38: 599-637. http://dx.doi.org/10.1080/10408699891274327
- Huslt AD, Verbeke N. Chiral separation by capillary [25] electrophoresis with oligosaccharides. J Chromatogr 1992; 608: 275-87. http://dx.doi.org/10.1016/0021-9673(92)87134-T
- Huslt AD, Verbeke N. Quantitation in chiral capillary [26] electrophoresis; theoretical and practical consideration. Electrophoresis 1994; 15: 854-63. http://dx.doi.org/10.1002/elps.11501501121
- [27] Huslt AD, Verbeke N, Carbohydrates as chiral selectors for capillary electrophoresis. Enantiomer 1997; 2: 69-79.
- Watanabe T, Takahasi K, Horiuchi M, Kato K, Nakazawa H, [28] Sugimoto T, Kanazawa H, Chiral separation and quantitation enantiomers by pentazocine Capillary of zone electrophoresis using maltodextrins. J Pharm Biomed Analysis 1999; 21: 75-81. http://dx.doi.org/10.1016/S0731-7085(99)00114-4
- Quang C, Khaledi MG. Direct separation of enantiomers of [29] beta-blockers by cyclodextrin-mediated capillary zone electrophoresis. J High Resolut Chromatogr 1994; 17: 609-12.

http://dx.doi.org/10.1002/jhrc.1240170810

- [30] Ozoemena KI, Stefan RI, van Staden JF, Aboul-Enein HY, Utilization of maltodextrin based enantioselective, potentiometric membrane electrodes for the enantioselective assay of S-perindopril. Talanta 2004; 62: 681-5. http://dx.doi.org/10.1016/j talanta.2003.08.035
- [31] Nejem RM, Stefan RI, van Staden JF, Aboul-Enein HY, Enantioanalysis of L-hydroxyglutaric acid in urine samples using enantioselective, potentiometric membrane electrodes based on maltodextrins. Talanta 2005 65: 437-0. http://dx.doi.org/10.1016/j.talanta.2004.06.040
- Stefan RI, Nejem RM, Enantioanalysis of glyceric acid in [32] urine samples using enantioselective, potentiometric membrane electrodes based on maltodextrins. Sens Actuators B 2005 106: 736-0. http://dx.doi.org/10.1016/j.snb.2004.09.031
- Stefan-van Staden RI, Bokretsion RG, van Staden JF, Aboul-[33] Enein HY, Enantioanalysis of butaclamol using enantioselective, potentiometric electrodes. Anal Lett 2009 42: 1111-8. http://dx.doi.org/10.1080/00032710902890462

Wootton IDP. Micro-analysis in Medical Biochemistry (4th [34] Edition), London: J.A. Chuchill Ltd. 1964.